

Limnology 08-15 Macrophytes

Aquatic Plants

Submersed Aquatic Vegetation (SAV)

Frog moss or moss

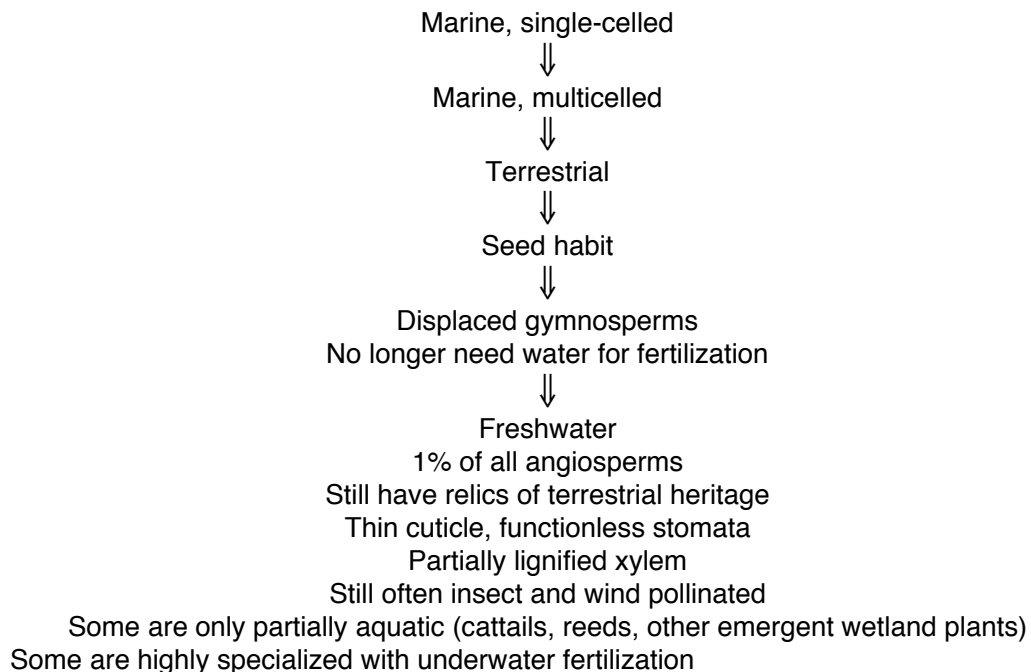
Seaweed

- Common taxa
- Morphological and biochemical adaptations to life underwater
- Ecological function
- Invasive species characteristics
- Management

Table 4.1. Annual production levels of fringing benthic plant communities. Average levels are compiled from all available data and represent a range of environments though data from very unfavourable sites is probably not included. The maximum values are the highest that could be located. The data are given in g organic matter (ash-free dry wt.) per m² per year.

Fringing community type	Average production	Maximum production
Bog	900	1500
Marshes, freshwater		
<i>Typha</i> , cattail	2700	3700
<i>Carex</i> , sedge	1000	1700
<i>Phragmites</i> , reed	2100	3000
<i>Cyperus</i> , papyrus	7500	15000
Freshwater, tidal	1600	2100
Marshes, salt water tidal		
<i>Spartina</i>	2500	6000
Other grasses	1500	8500
<i>Salicornia</i>	3000	
Swamps, freshwater		
Bog, spruce	500	
Cedar, Cypress	4000	
Hardwood	1600	
Swamps, salt water		
Mangrove	3000	
Benthic microphytes		
Freshwater, lake	170	1560
Freshwater, spring	3000	3500
Marine	200	850
Submerged macrophytes		
Freshwater, temperate	650	1300
tropical		1700
Marine		
sea grasses, tropical	2000	8500
macroalgae, kelp	3400	4250
rockweeds	750	2100
green algae, tropical		4000
Floating, freshwater		
Duckweed	150	1500
Water hyacinth (<i>Eichornia</i>)		3300

Evolution of aquatic plants



Taxonomic classification

Macroalgae
Nitella (Nitella gracilis)
Stonewort or muskgrass (Chara vulgaris)
Taste and odor in drinking water/mosquito larvicide

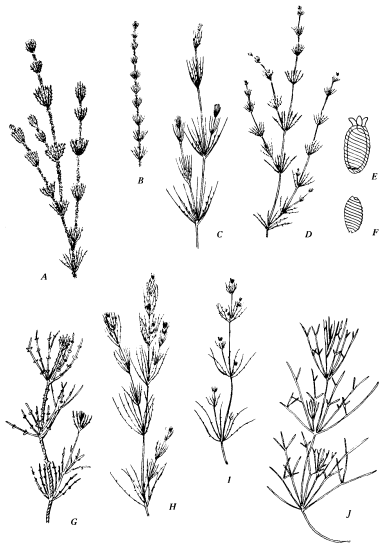


FIGURE 1. Typical members of the tribe Characeae of limnological interest. A, *Chara hispida*; B, *Ch. aspera*; C, *Ch. globularis*; D, *Ch. contraria*; E, intact oogonium of same showing three of the five cells of the coronula; F, the same as ordinarily fossilized; G, *Ch. tomentosa*; H, *Ch. radix*; I, *Ch. demodata*; J, *Nitellopsis obtusa* (from British specimens, Groves and Bullock-Webster).

Source: Hutchinson 1975

TABLE 3. Survivorship of *Aedes* larvae in water in which various *Charophyceae* are growing

Species of Charophyceae	Initial no. of <i>Aedes</i> larvae	Number after 30 hr	Number after 60 hr	Number after 120 hr
<i>Chara globularis</i>	10	10	10	9
<i>Ch. corallina</i>	10	8	6	2
<i>Ch. zeylanica</i>	10	9	5	4
<i>Nitella flexilis</i>	10	0	0	0
<i>N. rigida</i>	10	2	0	0
<i>N. mucronata</i>	10	4	1	1
Control, no plants	10	10	9	8

Source: Hutchinson 1975

Bryophytes

Liverworts

Riccia spp. primarily in eutrophic lakes – Corps reservoir

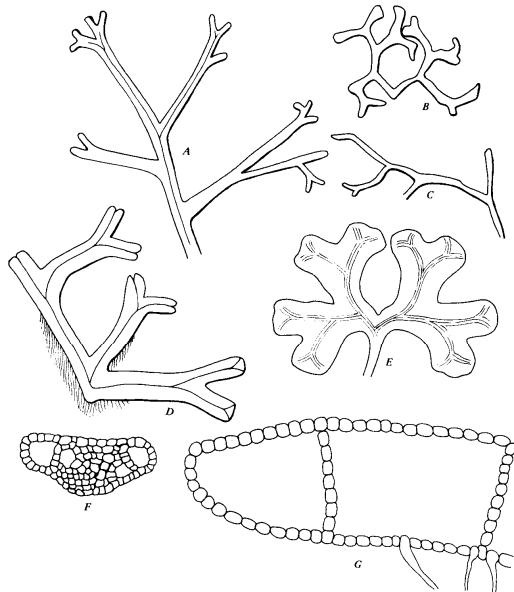


FIGURE 22. A, *Riccia fluitans* typical aquatic form; B, C, *R. rhenana*, typical aquatic forms, from Munich; D, *R. fluitans* terrestrial form; E, part of B cultivated terrestrially; F, cross-section of part of C; G, cross-section of part of E (A-E, $\times 3.75$; F, G, $\times 7.5$, Watson, Müller).

Source: Hutchinson 1975

Mosses

Order: Sphagnales.

Typically in calcium-poor habitats ranging from damp clearings in woodlands to littoral zone of lakes. Many species with distinct zonation



FIGURE 24. Aquatic mosses. A, *Sphagnum cuspidatum*; B, *S. cuspidatum* f. *plumosum*; C, *Scorpidium scorpioides*; D, *Fissidens julianus*; E, *F. grandifrons*; F, *Earhynchium rusciforme*; G, *Fontinalis antipyretica* (x1); H, *F. dalecarlica*; I, *Drepanocladus fluitans*; J, *Amblystegium riparium* (all natural size except G; Paul and Monkenmeyer slightly modified, all presumably based on European material).

Source: Hutchinson 1975

Order: Bryales

Ordinary mosses can be a major component of lake flora and production – very common in Corps of Engineers reservoirs surveyed in 1999 (*Fontinalis antipyretica*)

Mosses and liverworts can be a major component of the flora in oligotrophic lakes. Mosses and liverworts can grow deeper than angiosperms, which are limited by pressure to depth less than 11 m. Mosses and liverworts can grow as deep as 164 m, e.g., Waldo Lake, Crater Lake, Tahoe.

Major growth forms

Pleustophytes: free floating

Rhizophytes: rooted in the sediment

A general classification scheme

Emergent – stems and leaves held above the water surface, usually rooted. Found on exposed soil at lake margin to 150 cm.

Purple loosestrife (*Lythrum salicaria*)

Cattails (*Typha latifolia*)

Common reed (*Phragmites australis*)

Submersed – stems and leaves entirely underwater (may have emergent flowers), rooted in the sediment.

Eurasian watermilfoil (*Myriophyllum spicatum*)

Brazilian elodea (*Egeria densa*)

Narrow-leaf pondweed (*P. pusillus*)

Floating – leaves on the water surface, submersed leaves may also be present, rooted in the sediment.

Water lilies (*Nuphar polysepalum*, *Nymphaea alba*)

American pondweed (*Potamogeton nodosus*)

Big-leaf pondweed (*P. natans*)

Water shield (*Brasenia schreberi*)

Free-floating – plants not rooted in the sediment

With emergent or floating leaves

Water hyacinth (*Eichhornia crassipes*)

Duckweed (*Lemna minor*, *L. trisulca*)

With submersed leaves

Coontail (*Ceratophyllum demersum*)

Bladderwort (*Utricularia vulgaris*)

General classification schemes, by their nature, cannot capture all the potential variations in growth form, but they do provide a quick reference to ecology and function of the plants in the ecosystem

Examples of problem plants in this classification scheme

Parrotfeather (*Myriophyllum aquaticum*) – submersed in winter, emergent in spring, free-floating in summer

Water hyacinth (*Eichhornia crassipes*) – seedlings germinate on exposed mudflats, remain rooted in shallow water, become free-floating in deep water

Emergent

Typha latifolia
Broadleaved cattail



Iris pseudacorus
Yellow flag iris

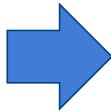


Lythrum salicaria
Purple loosestrife



Floating leaf, rooted

Nymphaea odorata



Nuphar polysepalum



Floating leaf, rooted *Potamogeton natans*

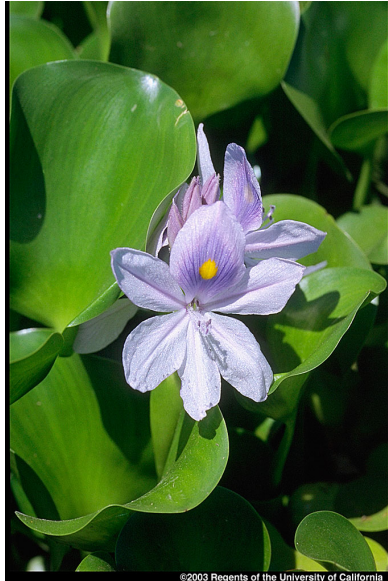


Submersed – rooted *Hydrilla verticillata*



Floating leaf-unrooted
in water column

Water hyacinth
(*Eichhornia crassipes*)

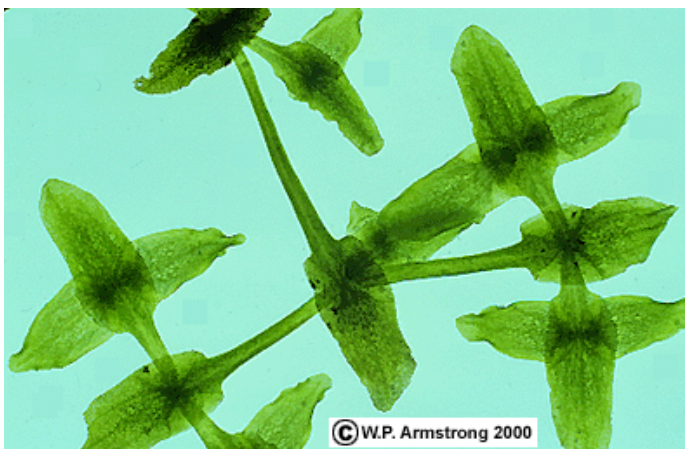


Submersed – not rooted

Ceratophyllum demersum



Lemna trisulca



Floating – not rooted in sediment

Ferns

Azolla (Water velvet) – floating, red thallus that contains symbiotic cyanobacteria *Anabaena azollae*, a nitrogen-fixing organism.

Salvinia (Water fern) – *S. molesta*, *S. minima* are introduced, weedy species

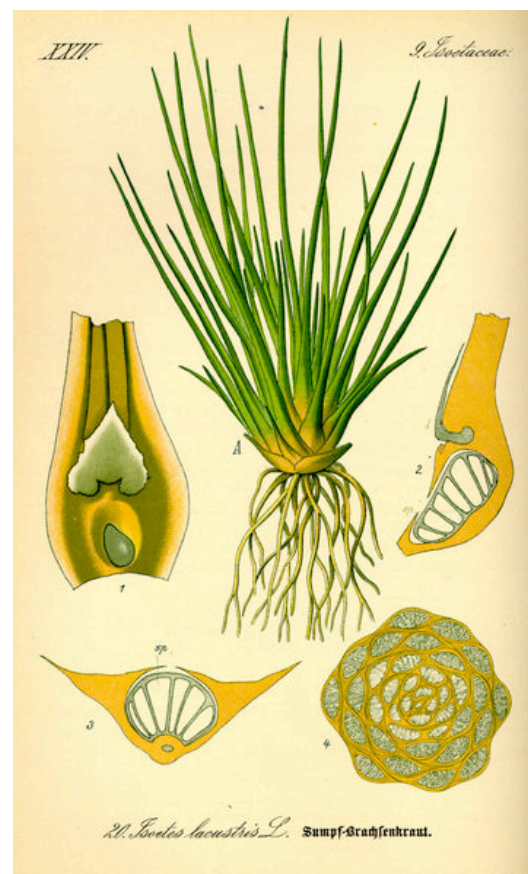


Angiosperm

Lemna spp (duckweeds)

Isoetids - “fern allies”

Typical in oligotrophic lakes,
evergreen, slow-growing,
long-lived leaves



Angiosperms (Vacular hydrophytes)

Aquatic angiosperm flora is enriched in monocots relative to terrestrial flora

monocot;dicot ratio

1:1 in aquatic flora

1: 4–5 in terrestrial flora

Many families, but few species/ family

Monocots

Hydrocharitaceae

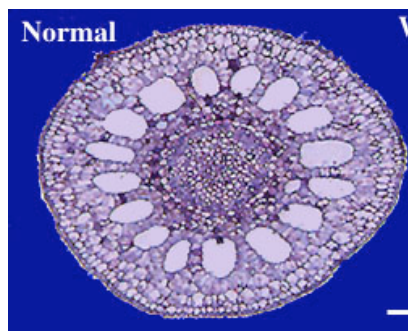
Egeria densa

Elodea canadensis

Hydrilla verticillata

Environmental influences on aquatic plant growth

- Density
 - Buoyancy due to aerenchyma tissue
 - Reduced requirement for structural tissues

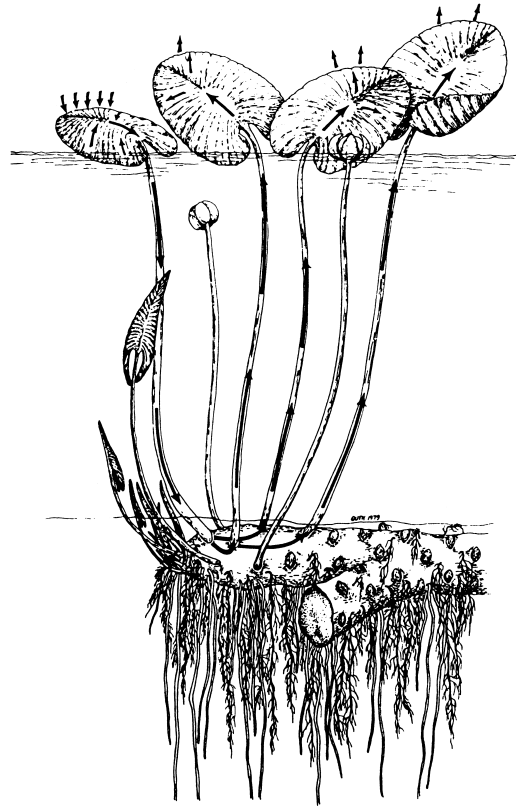


xs of *Myriophyllum spicatum* leaf

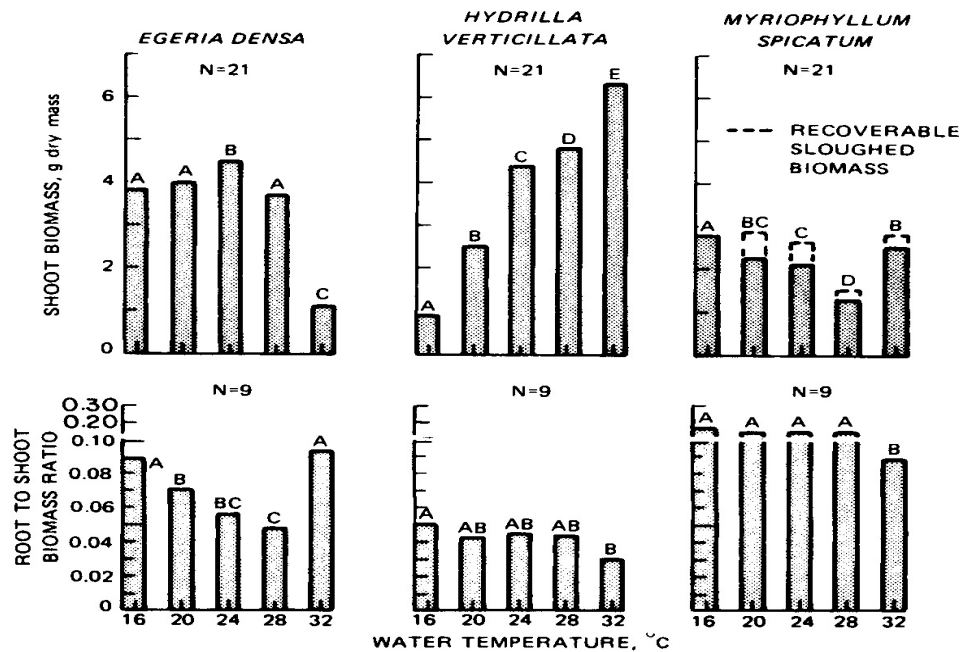
Aerenchyma tissue allows the transport of gasses to roots and rhizomes in anaerobic sediments

Pressurized ventilation in water lilies is driven by counterbalancing Leaf temperature and internal water vapor effects on diffusion of gases from the dryer atmosphere

See Dacey, 1981, Ecology 62: 1137-1147

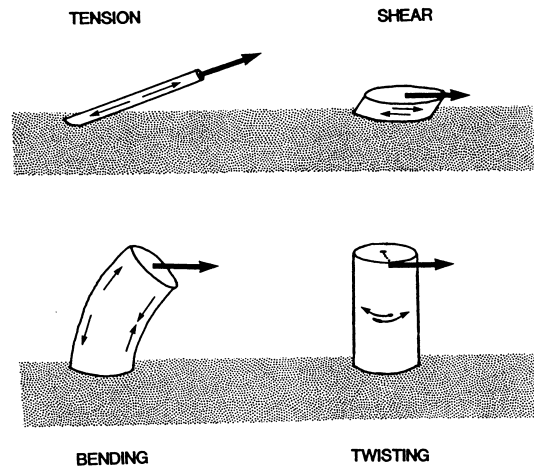


Growth and allocation of biomass influenced by temperature
(species dependent)



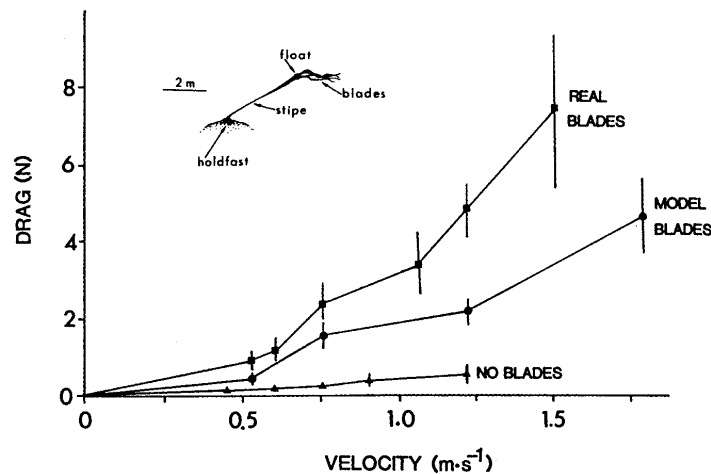
- Viscosity effects

–Submersed aquatic plants are subject to tensional stresses while terrestrial plants are subject to compressional stresses due to gravity



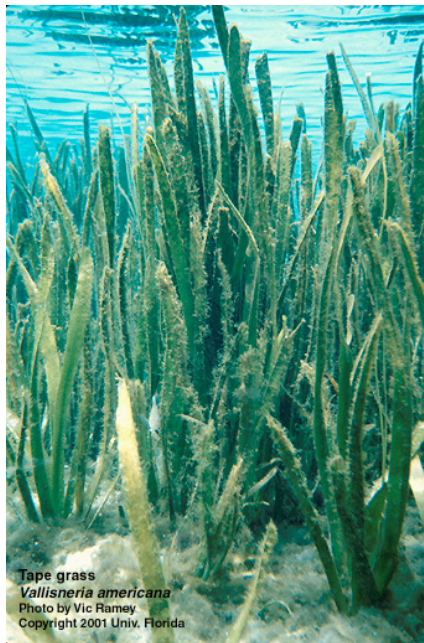
Stresses caused by water viscosity on sessile organisms

Source: Vogel. Life in Moving Fluids.



Drag force measured on a *Nereocystis leuteana* (squares), on a stipe and pneumatocyst (float) after the fronds had been removed (triangles) and on the stipe an pneumatocyst plus plastic fronds (no ruffles as on blades).
Source: Koehl.

Strap-like leaves of *Vallisneria americana* and *Stuckenia pectinata* reduce drag in high-flow environments



Dissected leaves of eurasian watermifoil (*Myriophyllum spicatum* and *Cabomba caroliniana*) limit distribution to low-flow environments



–Gas diffusion is slow in water

$$D_{\text{CO}_2} \text{ in air} = 1.47 \times 10^{-5} \text{ m}^2/\text{s}$$

$$D_{\text{CO}_2} \text{ in water} = 1.8 \times 10^{-9} \text{ m}^2/\text{s}$$

$$D_{\text{O}_2} \text{ in air} = 0.188 \times 10^{-4} \text{ m}^2/\text{s}$$

$$D_{\text{O}_2} \text{ in water} = 2 \times 10^{-10} \text{ to } 1.2 \times 10^{-9} \text{ m}^2/\text{s}$$

Low diffusion rates result in CO₂ depletion and O₂ accumulation near leaves during photosynthesis, with consequences for photorespiration and carbon-fixation rates

Adaptations to reduce boundary layer effect

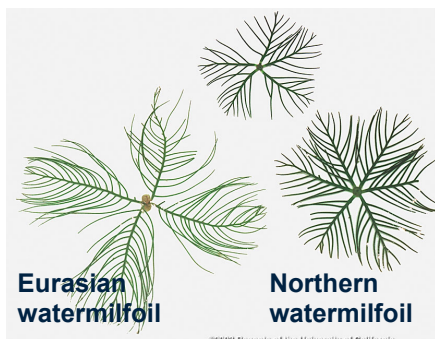
Increase surface to volume ratio

Thin and dissected leaves

Wavy leaves

Emergent leaves

Biochemical



Dissected, thin, and narrow leaves result in high surface to volume ratio, which maximizes surface available for gas exchange

Eurasian watermilfoil
Myriophyllum spicatum



Sago pondweed
Stuckenia pectinatus

Curlyleaf pondweed
Potamogeton crispus

Wavey leaves break up
boundary layer



Heterophylly is an adaptation to limits on gas diffusion

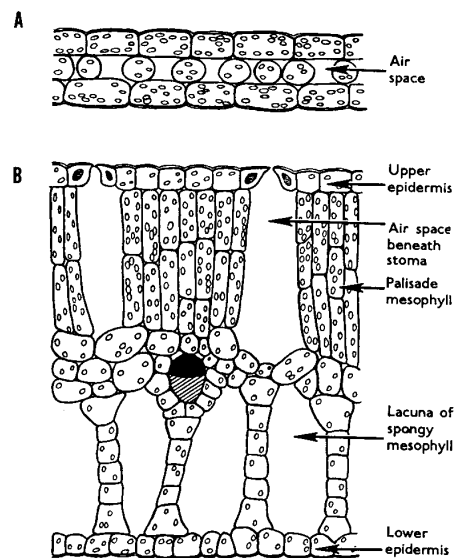
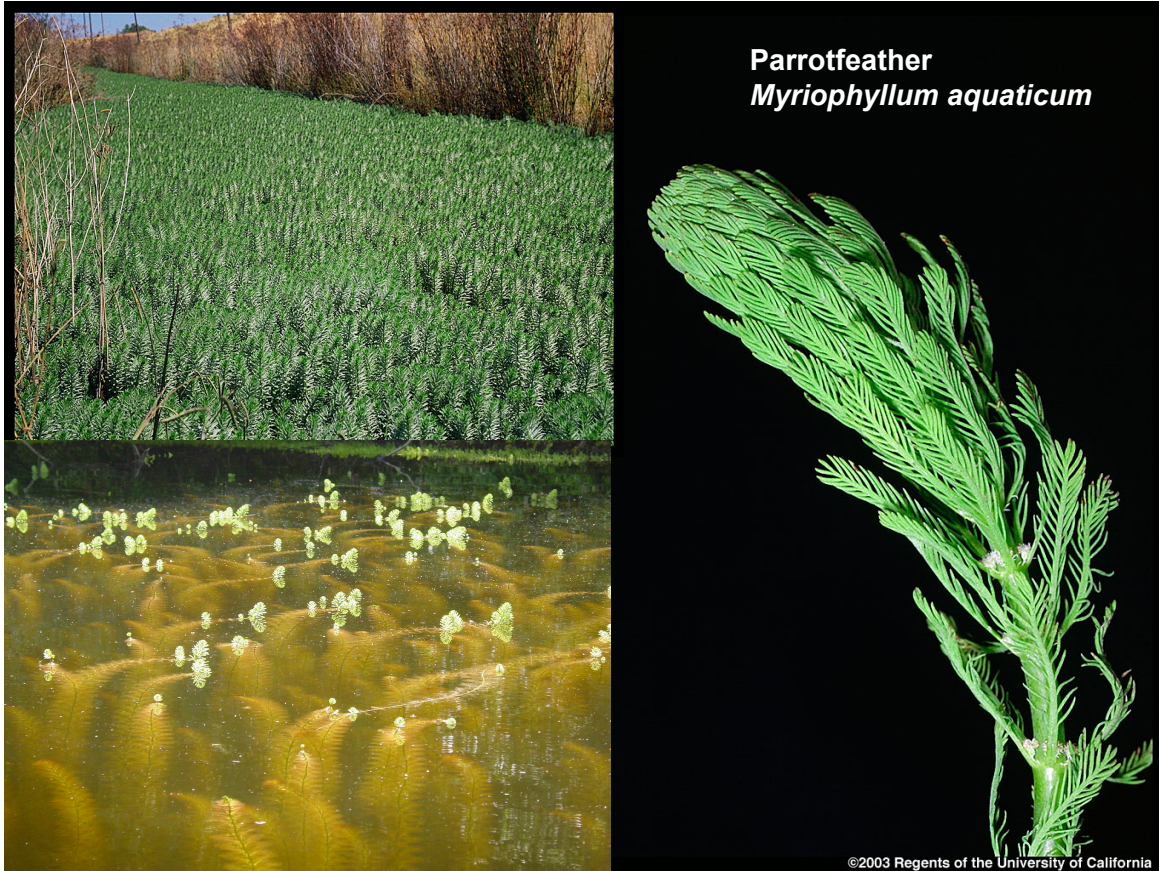


FIG. 8.10. *Potamogeton nodosus*: T.S. parts of submerged ribbon leaf (A) and floating leaf-blade (B), showing contrasting anatomy. Note reduction of submerged leaf to three cell layers with abundant epidermal chloroplasts: floating leaf with stomata in the upper epidermis, few epidermal chloroplasts, and well differentiated mesophyll; xylem of vascular bundle in solid black, phloem shaded (A, $\times 240$; B, $\times 180$) (after Streitberg, 1954).

Source: Sculthorpe



- Effect of Light attenuation

- Submersed plants are "shade plants", net photosynthesis is saturated at $< 1/2$ full-sun
- Low photosynthesis rates - light and CO₂ saturated photosyn. rates rarely exceed 100 $\mu\text{mol/mg Chla/hr}$
- At naturally occurring DIC conc. Rates are closer to 10-20 $\mu\text{mol/mg Chla/hr}$
- pH effects on DIC availability may be more important than light in determining PS rates