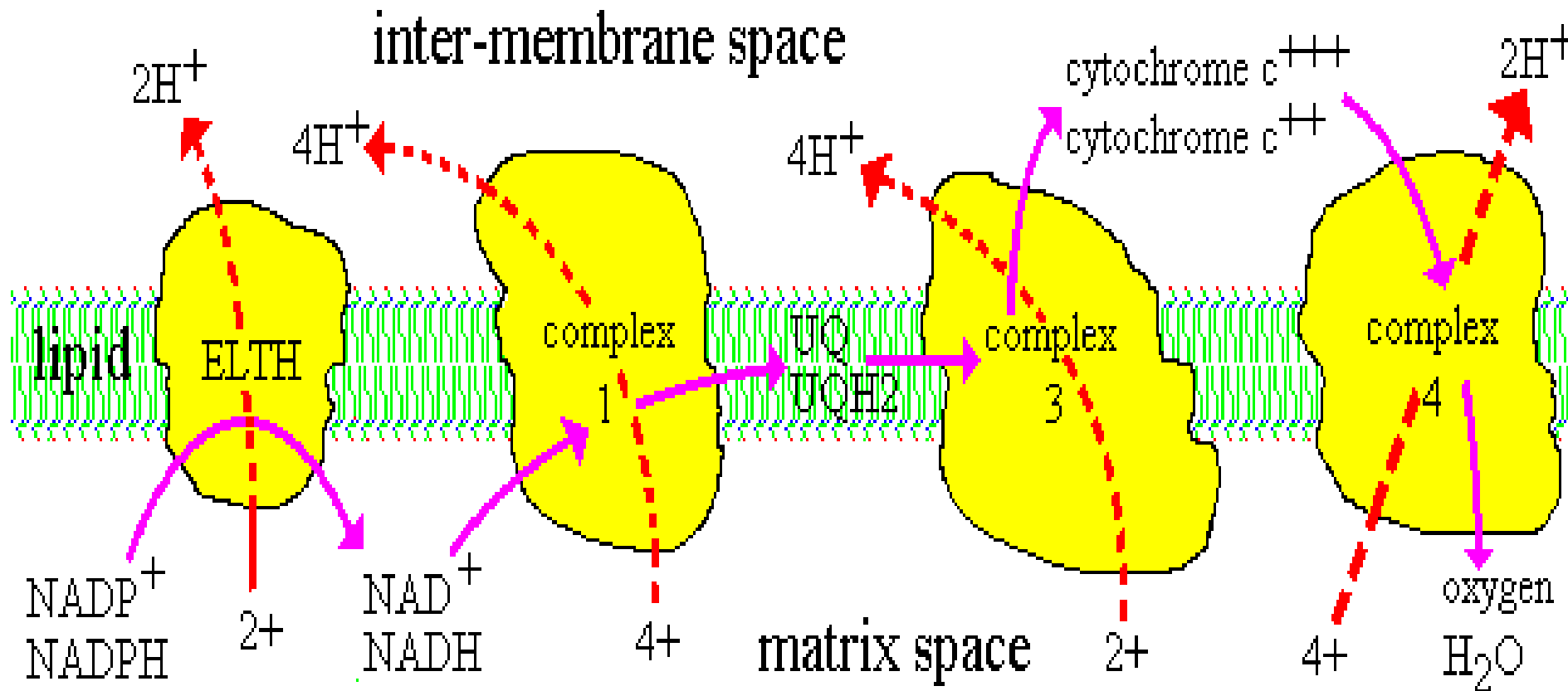


The Q-Cycle

A contribution to Dr. Keith Garlid's
Bioenergetics course on the proton-
translocating bc_1 complex (Complex III)
by Ian West (6th May 2004)

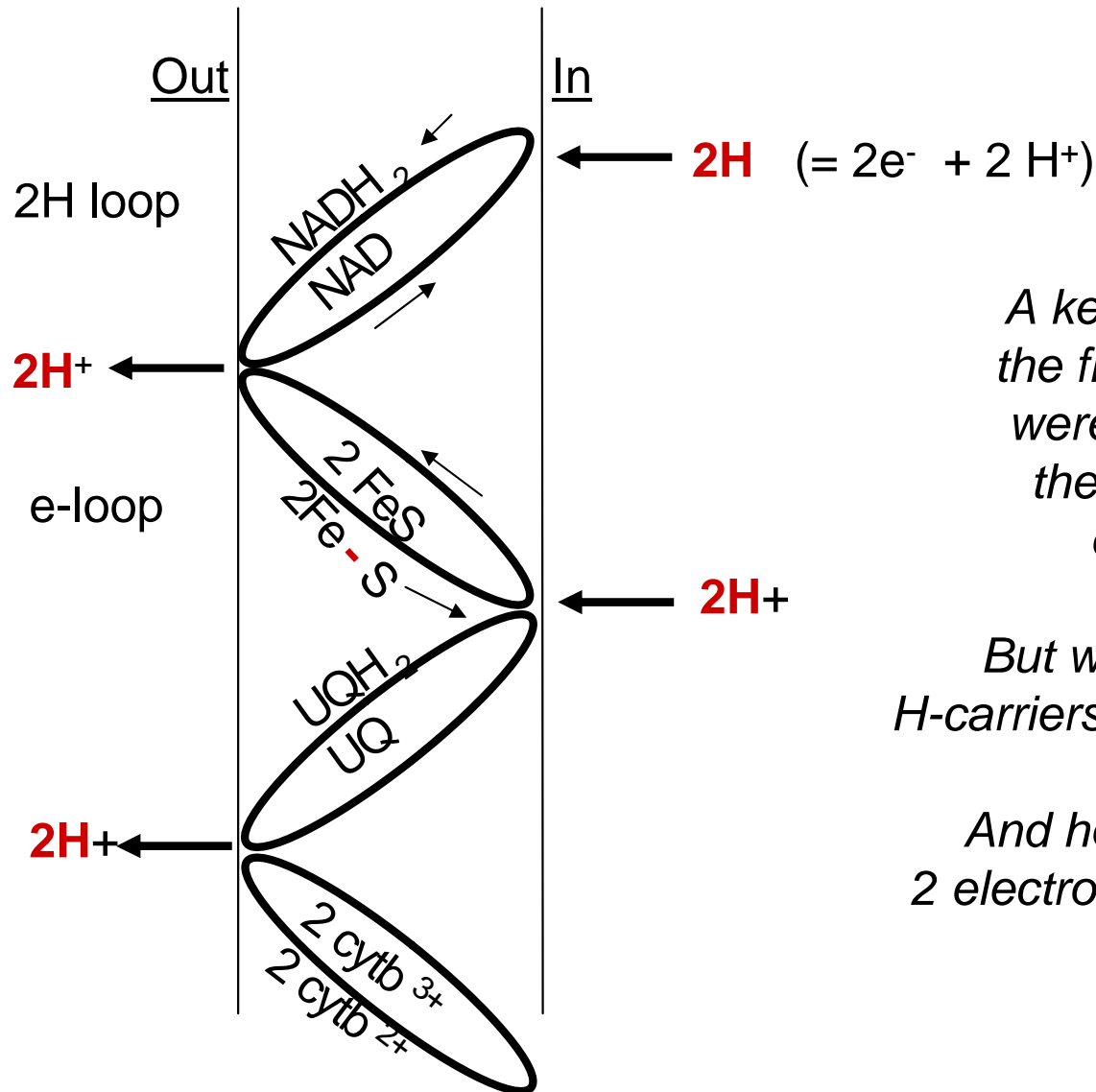
Handy web addresses

- http://www.life.uiuc.edu/crofts/bc-complex_site/
- <http://www.life.uiuc.edu/crofts/bioph354/index.html>
- <http://www.bmb.leeds.ac.uk/illingworth/oxphos/index.htm>
- <http://www.cmb.uab.edu/courses/Lectures/brooks2.pdf>
- <http://fig.cox.miami.edu/~cmallery/255/255etc/CoQ.jpg>
- www.medphys.ucl.ac.uk/research/borl/research/IR_topics/spectra/spectra.htm



Mitchell's suggestion of how electron flow might be coupled to proton-translocation

"Direct-Coupling"



A key idea was that the flow of e^- and H^+ were co-linear, and the stoichiometry one-to-one.

But what could be the H-carriers, and the e-carriers?

And how can cyt b take 2 electrons from e.g. UQH_2 ?

Redox Components

FMN (2e, 2 H⁺)?

UQ₁₀ (2e, 2 H⁺)?

FeS2 FeS1
FeS3

FeS4
FeS5
FeS6
FeS7
FeS8
FeS9

NADH⁺ + H⁺
(2e, 1 H⁺)

Cyt b -- cyt **b_L**
 cyt **b_H**
Cyt c -- cyt **c₁**
 cyt **c₁**
Cyt a -- cyt **a**
 cyt **a₃**

} (1e)

} (1e)

What is a redox midpoint potential ?



$$\text{pH} = \text{pK}_A + \text{Log} (\text{CH}_3\text{COO}^- / \text{CH}_3\text{COOH})$$



$$E_h = E_0' + \frac{2.3RT}{nF} \text{Log} (\text{Cyt c(III)}^{3+} / \text{Cyt c(II)}^{2+})$$

So, E_h is the “activity” of the electron relative to a standard which is that of electrons in a mix of H_2 gas and 1 M H^+

While E_0' is a constant that reflects the affinity of X^+ for e^-

Succinate ($E_0' = +30$ mV) can give electrons to UQ ($E_0' = +70$ mV) ; but not (spontaneously) to NAD^+ ($E_0' = -320$ mV)

UQH_2 cannot give one electron ($E_0' = +230$ mV) to cyt bL ($E_0' = -50$ mV), or even to cyt b_H ($E_0' = +50$ mV).

Redox Midpoints (E_0')

(all in mV)

FMN
- 220

FeS2
- 20

FeS1
- 220

FeS3

UQ₁₀ --- +70

NADH⁺ + H⁺
- 320

FeS4
FeS5
FeS6
FeS7
FeS8
FeS9

Cyt b -- cyt **b_L** --- - 50

cyt **b_H** --- +50

Cyt c -- cyt **c₁** --- +220

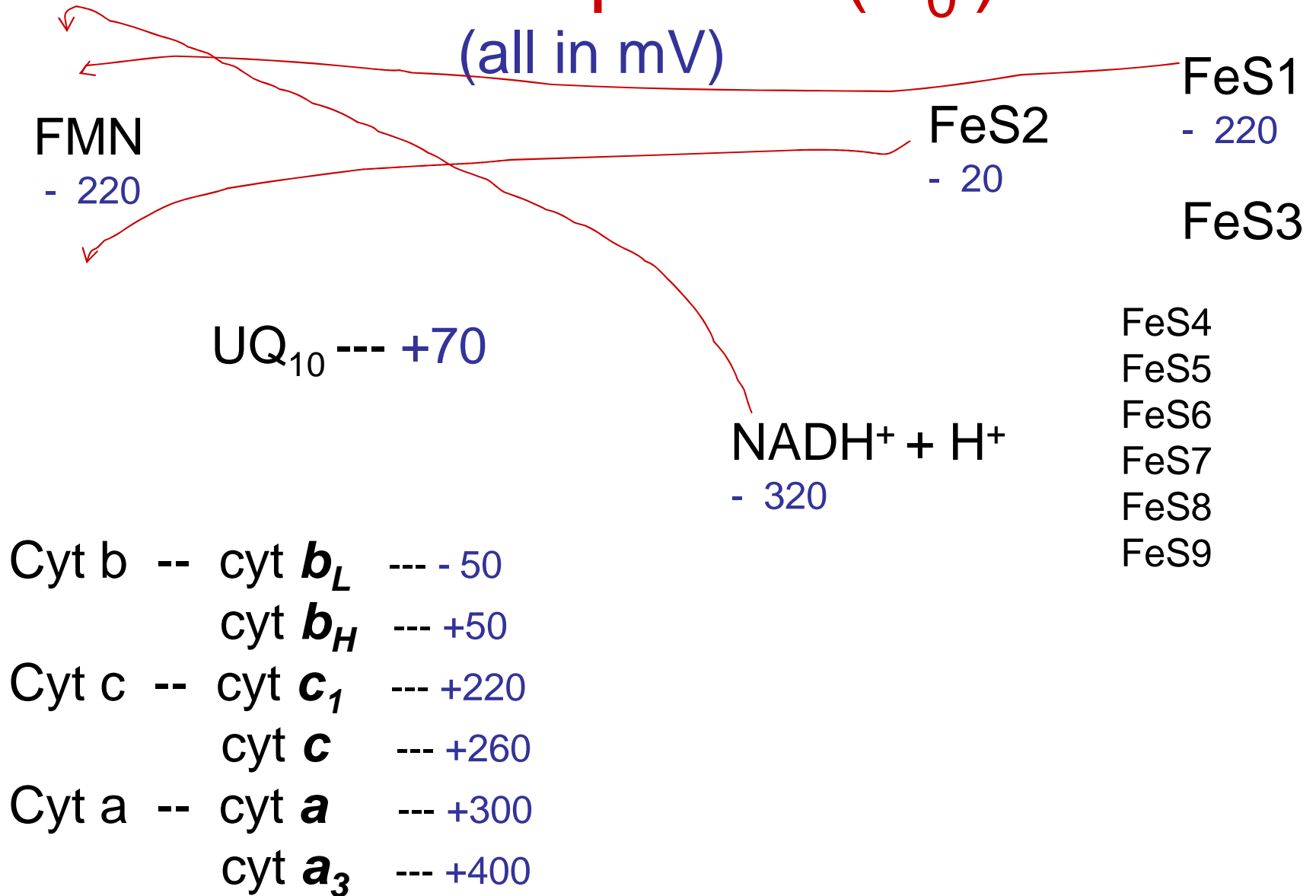
cyt **c** --- +260

Cyt a -- cyt **a** --- +300

cyt **a₃** --- +400

Redox Midpoints (E_0')

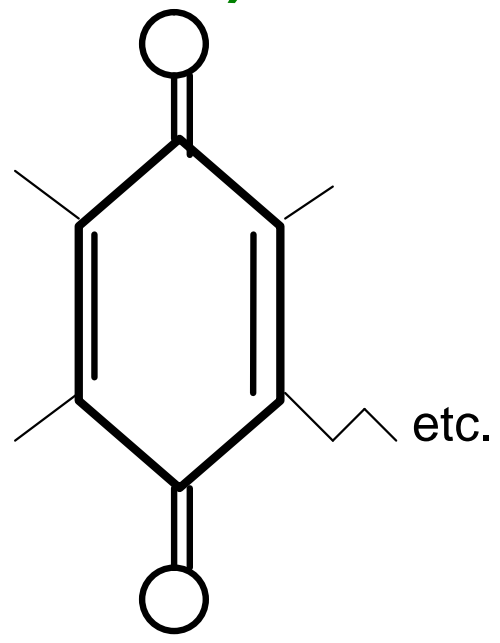
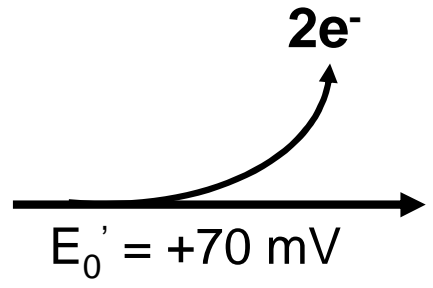
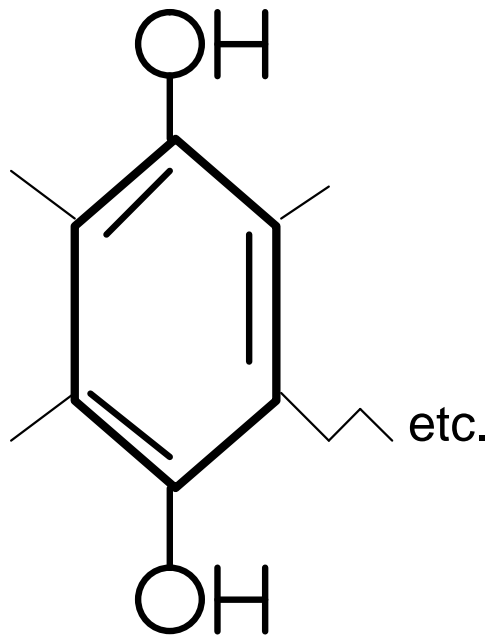
(all in mV)



UQ-negative mutants

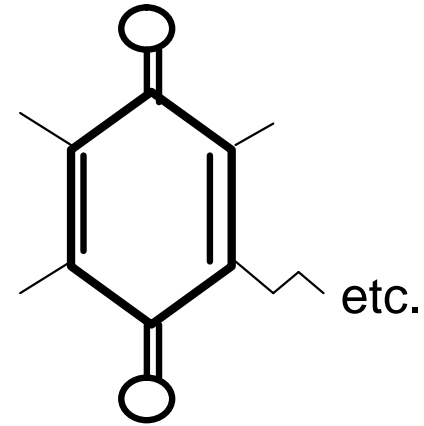
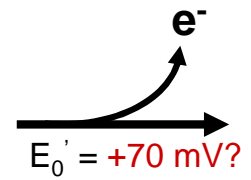
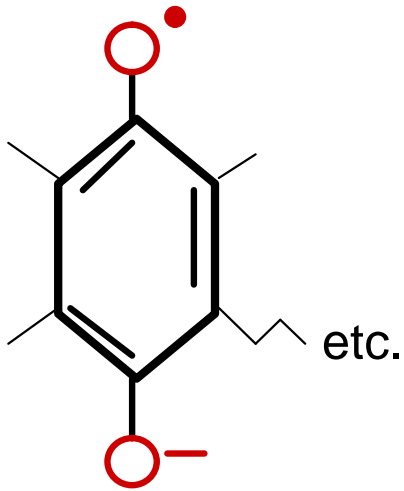
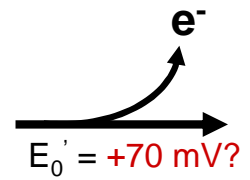
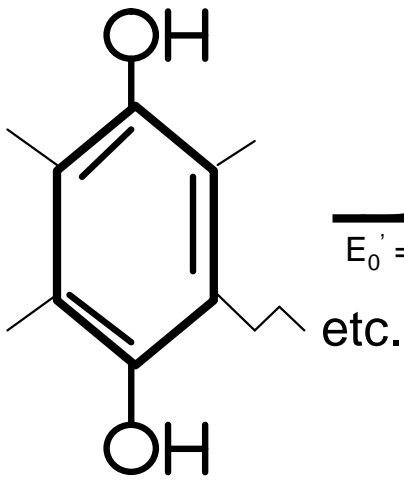
- In bacteria there are various cytochromes and quinones
- UQ-negative mutants cannot synthesize ubiquinone, or synthesize very little
 - Gibson F: The introduction of Escherichia coli and biochemical genetics to the study of oxidative phosphorylation. Trends.Biochem.Sci 25:342-344, 2000
 - Cox GB, Gibson F: Studies on electron transport and energy-linked reactions using mutants of Escherichia coli. Biochim.Biophys.Acta 346:1-25, 1974
- (I think) they showed that both the **reduction** and the **oxidation** of b-cytochrome(s) are slowed

UQ (ubiquinone)



H^+

H^+

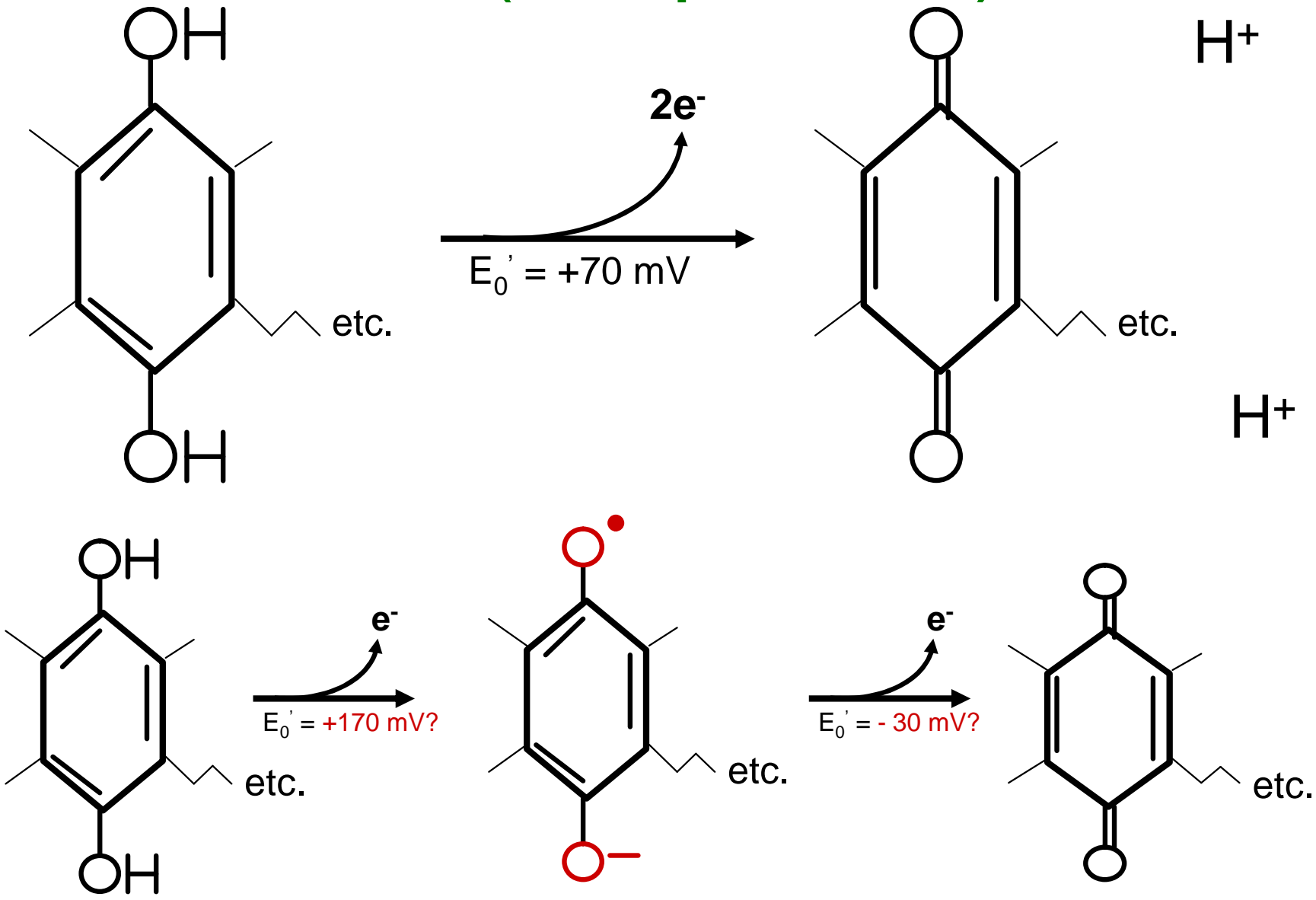


Ubiquinol

Semiquinone radical anion

Ubiquinone

UQ (ubiquinone)

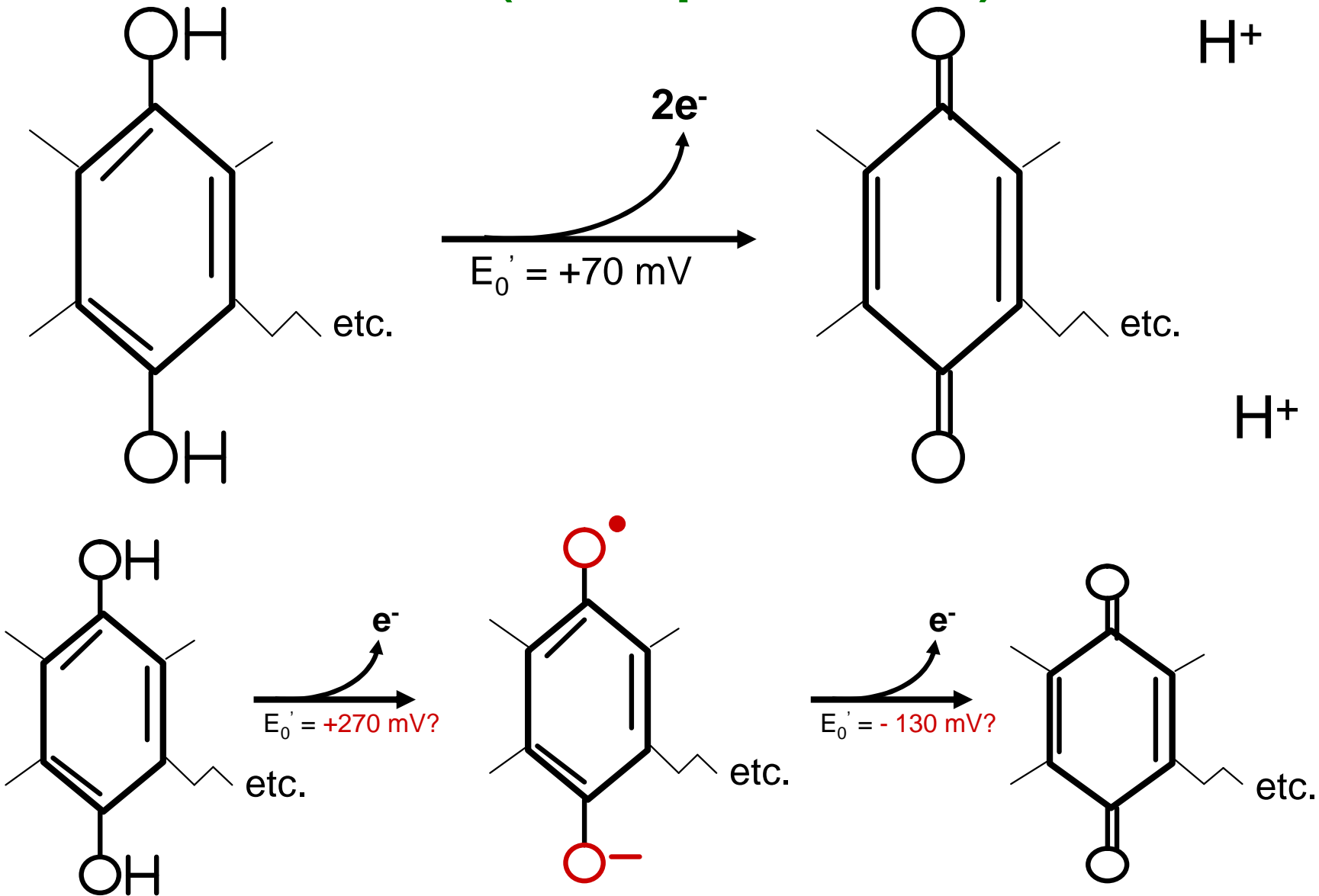


Ubiquinol

Semiquinone radical anion

Ubiquinone

UQ (ubiquinone)



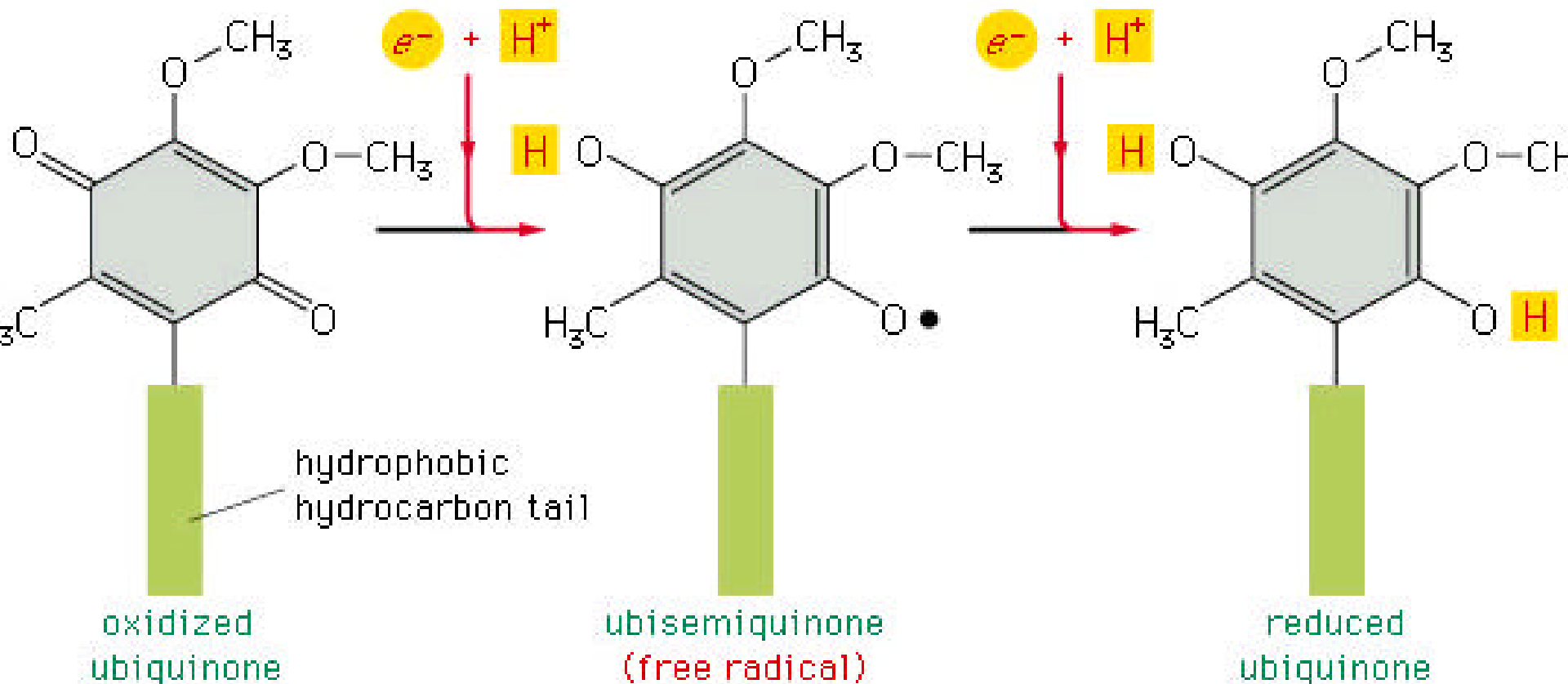
Ubiquinol

Semiquinone anion radical

Ubiquinone

Ubiquinone 10

- <http://fig.cox.miami.edu/~cmallery/255/255etc/CoQ.jpg>



Redox Midpoints (E_0')

(all in mV -- these are from memory!!)

FMN
- 220

FeS2
- 20

FeS1
- 220

FeS3

UQ₁₀ --- +70

NADH⁺ + H⁺
- 320

FeS4
FeS5
FeS6
FeS7
FeS8
FeS9

Cyt b -- cyt **b_L** --- - 50

cyt **b_H** --- +50

Cyt c -- cyt **c₁** --- +220

cyt **c** --- +260

Cyt a -- cyt **a** --- +300

cyt **a₃** --- +400

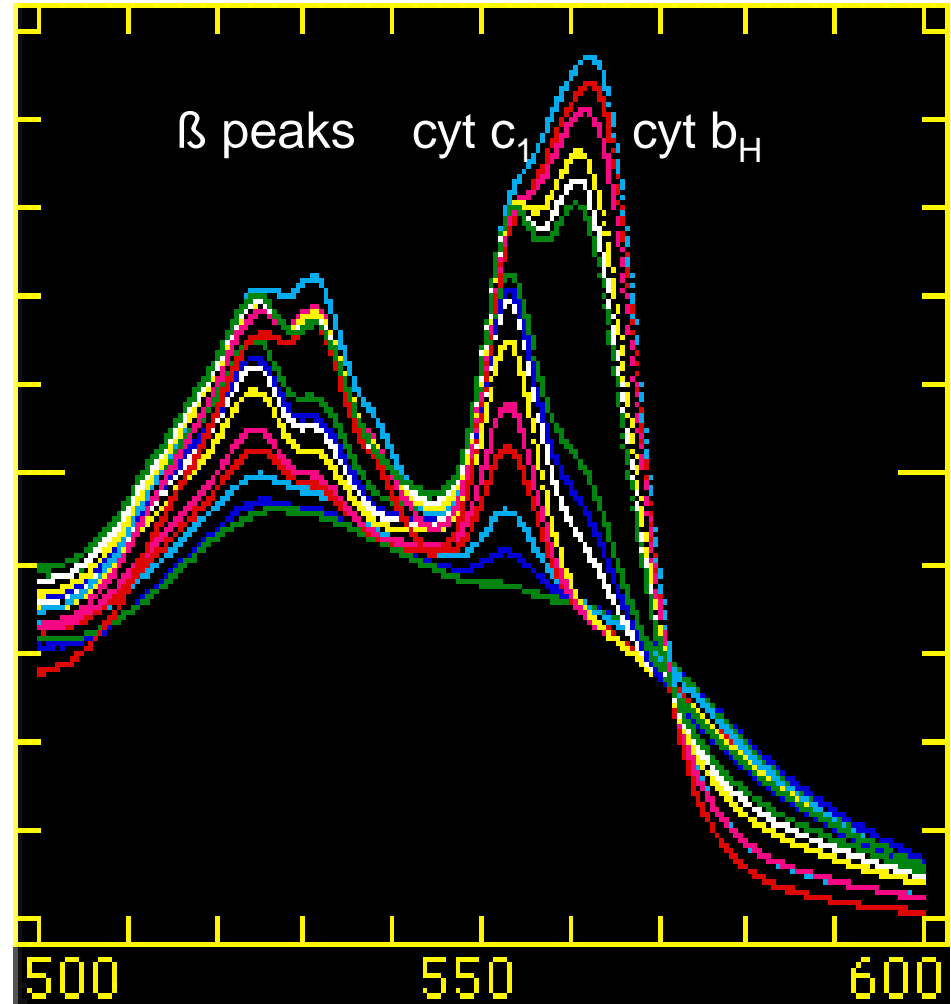
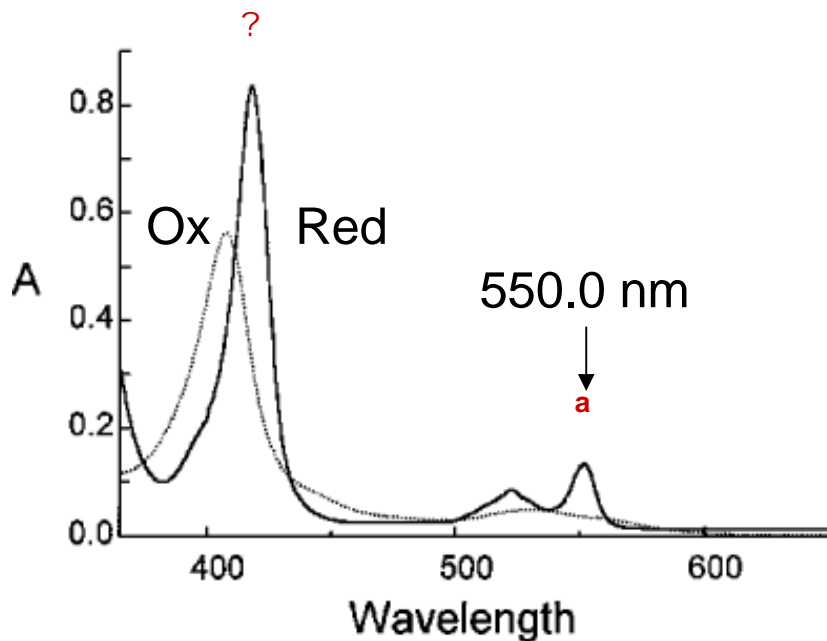
Fe(CN)₆(III)/(II) -- +430 (360)

O₂/ 2H₂O -- +820

Cytochrome spectra

Like litmus indicating pH, many compounds change colour on reduction

Cytochrome c spectra



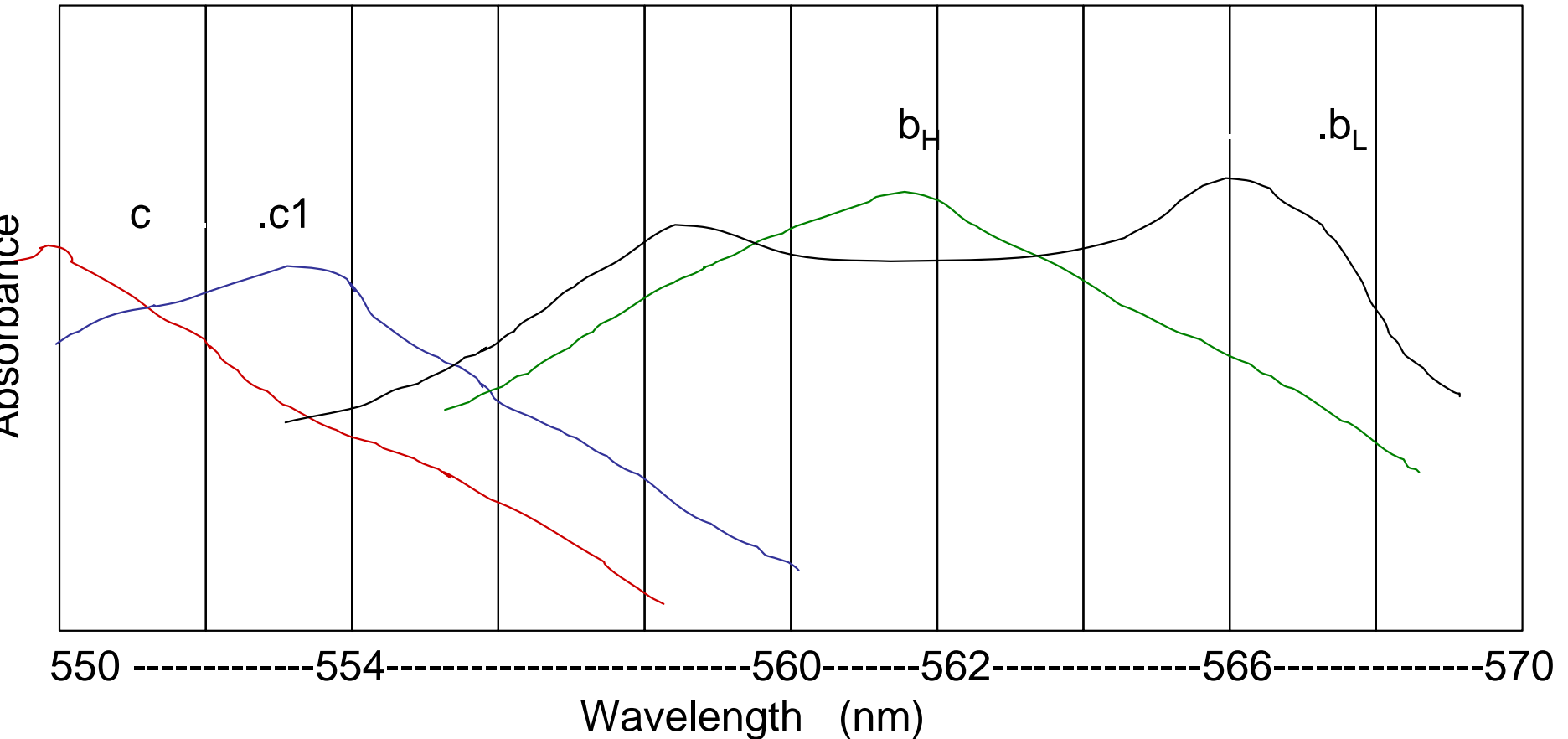
http://www.life.uiuc.edu/crofts/bc-complex_site/

Cytochrome spectra (2)

Like litmus indicating pH, many compounds change colour on reduction

This is a rough sketch of the alpha peaks only.

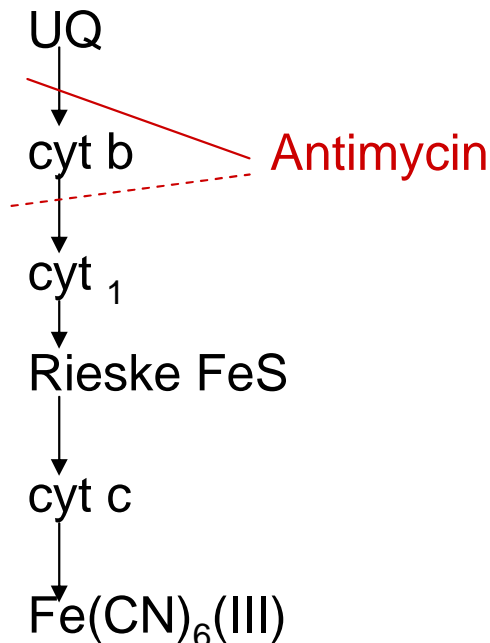
See West et al (1988) Biochim.Biophys.Acta 933:35-41.



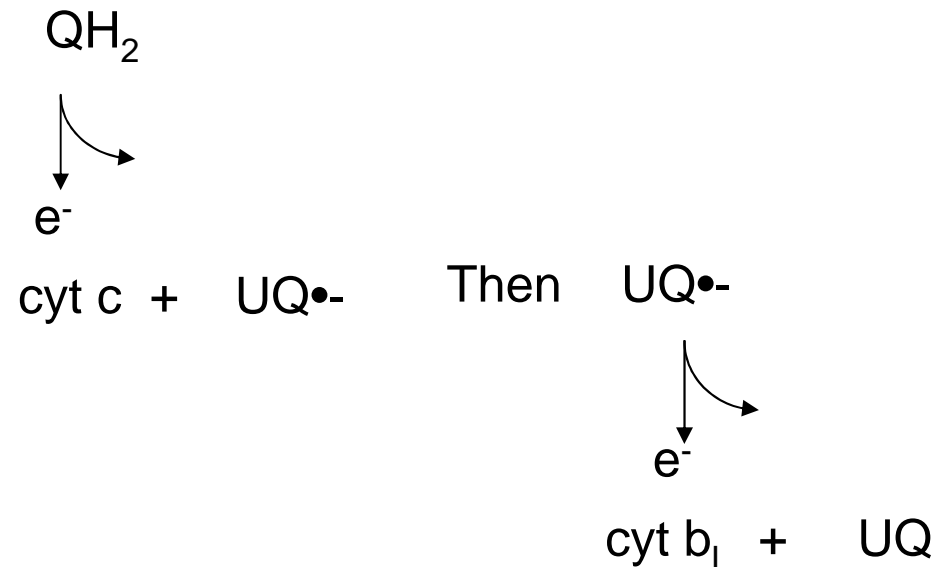
Wickström's see-saw

- Wickström (1972) showed that (in antimycin-inhibited mitochondria) adding an oxidant of cyt c caused reduction of cyt b_L ??

The Puzzle

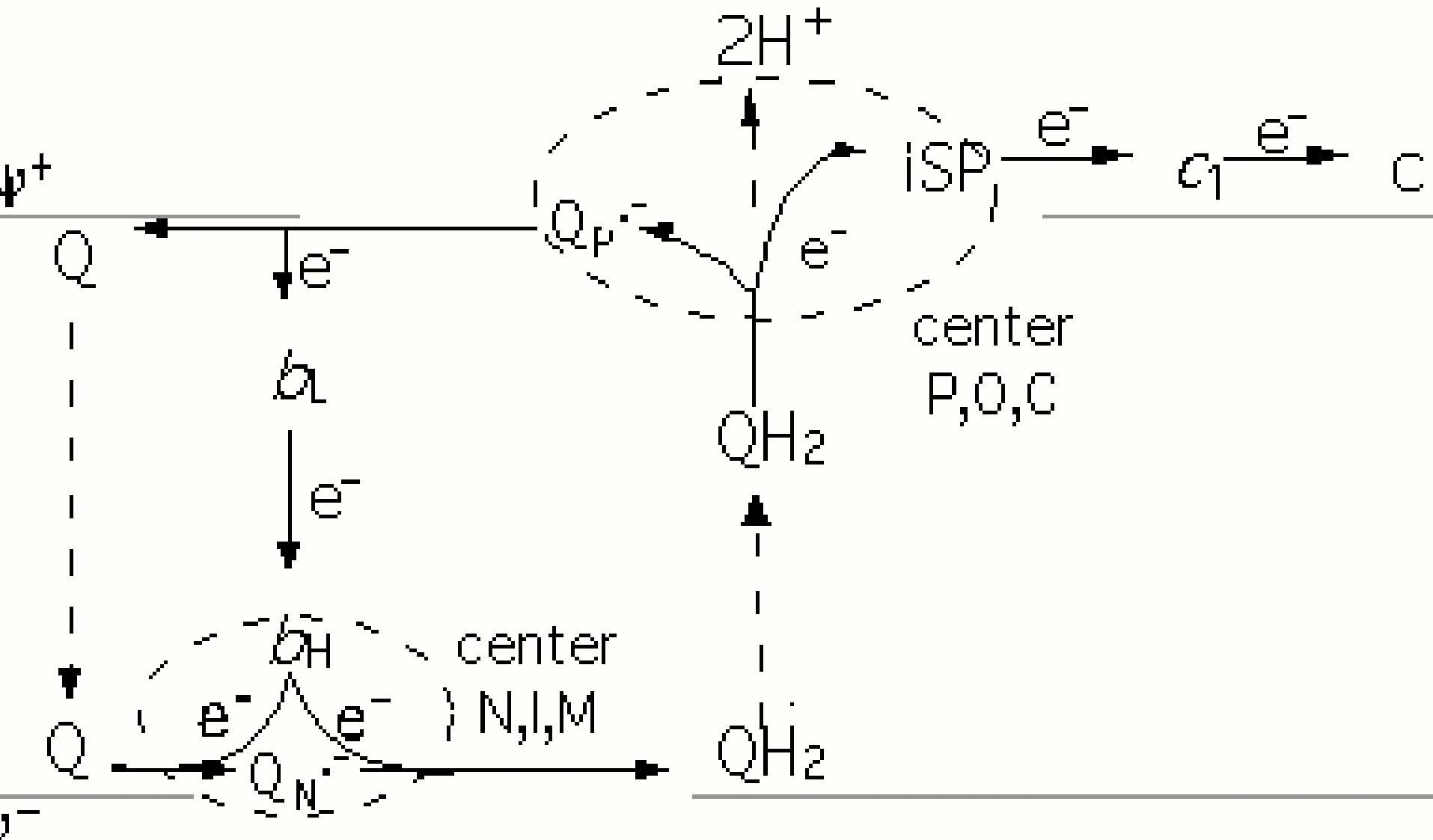


Wickström's inspired suggestion



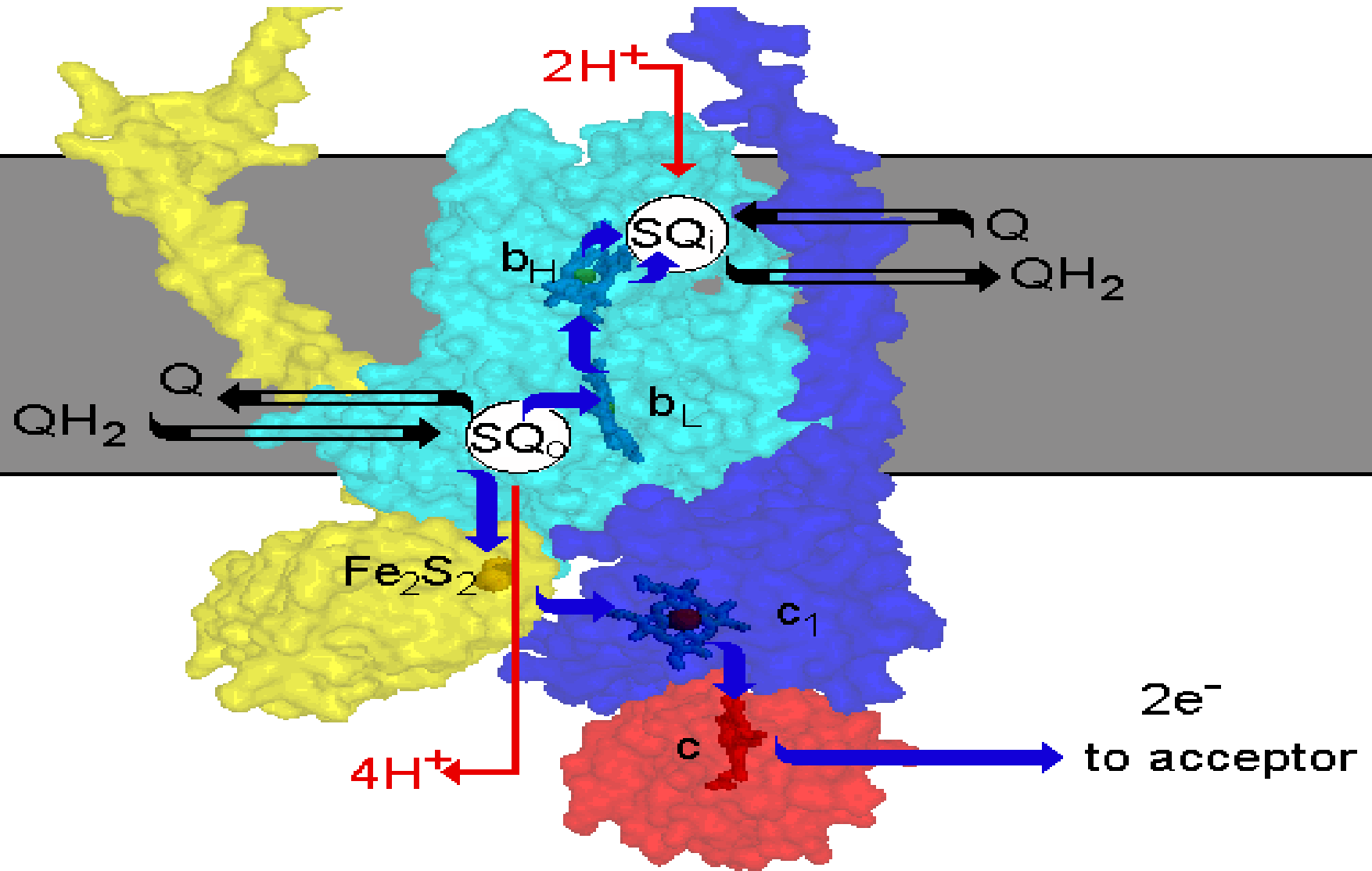
Mitchell's Q-cycle

(according to Yu Chang-An)



Mitchell's Q-cycle

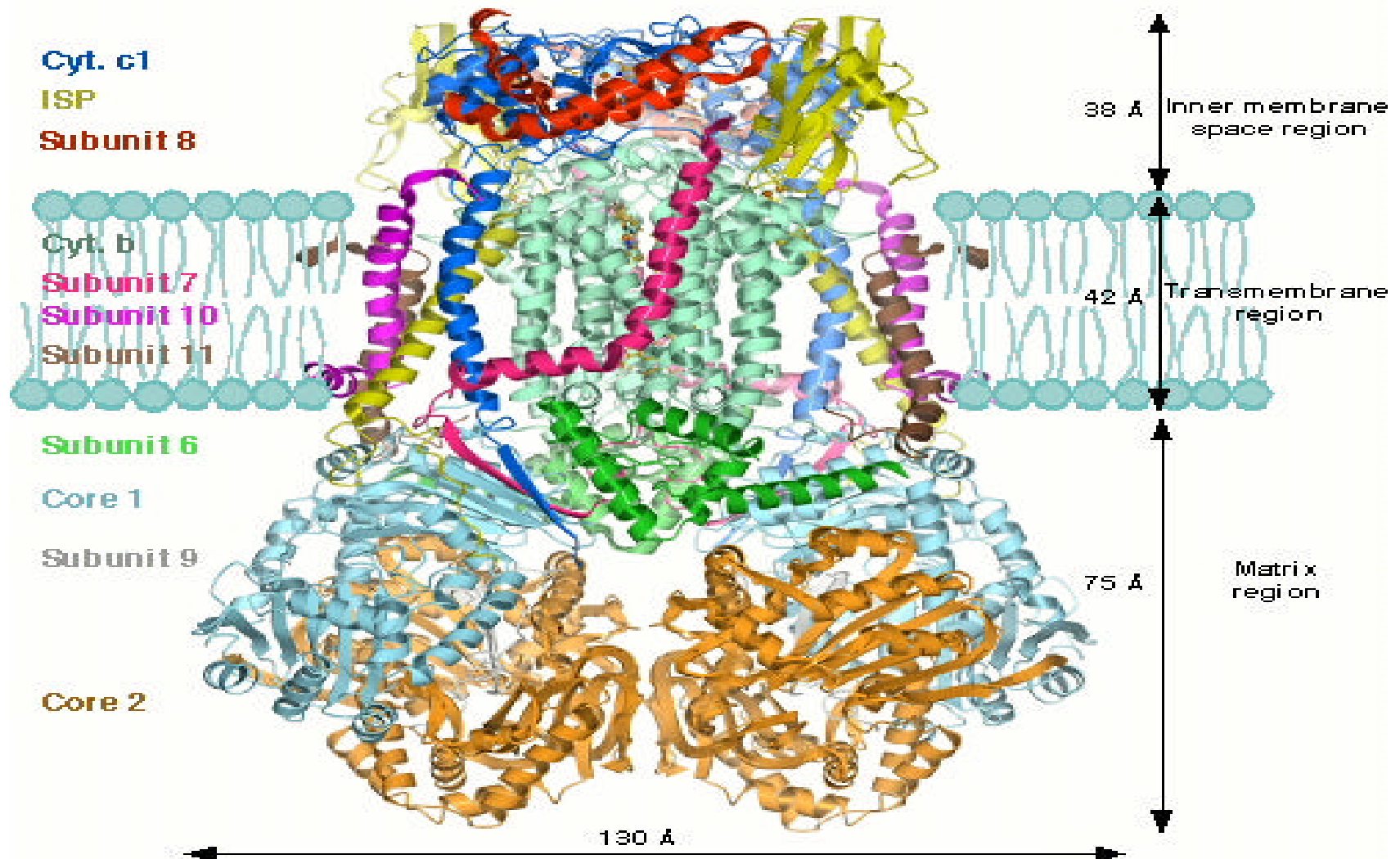
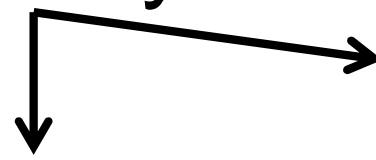
(According to Tony Crofts)



The overall stoichiometries

- The o-site turns over twice: 4 H⁺ to “o”
- The i-site turns over once: 2 H⁺ from “i”
- 2 UQH₂ are oxidized but 1 is reduced
- So, overall fuel consumption: 1 UQH₂
- 2 electrons flow to cyt c (and on to O₂)
- 2 electrons flow across membrane
(generating ??)
- Of the 4H⁺ to “o”, 2 come from “i” while 2
come from

'Google'-search "Q-cycle"



The bc1-complex from beef heart mitochondria

Subunit (no.)		Mr(beef)	Function
I	Core	53.6	No catalytic, protein transport Core
II	Core	46.5	No catalytic, protein transport
III	Cyt b	42.6	donor to Qi-site , acpt from SQ at Qo-site (8)
IV	c1	27.3	donor to cyt c (1)
V	Rieske's 2Fe.2S	21.6	acceptor from QoH2 , donor to cyt c1 Subunit (1)
VI	none	13.3	none known
VII	none	9.5	none known (1)
VIII	none	9.2	hinge protein (interacts with c1)
IX	none	8.0	none known
X	none	7.2	none known (1)
XI	none	6.4	none known (not present in chicken)

A.R. Crofts' Q-cycle web addresses

- <http://www.life.uiuc.edu/crofts/bioph354/index.html>
- http://www.life.uiuc.edu/crofts/bioph354/bc-complex_summary.html
- http://www.life.uiuc.edu/crofts/bc-complex_site/
- http://www.life.uiuc.edu/crofts/bc1_in_chime/bcc.html
- http://www.life.uiuc.edu/crofts/bc-complex_site/etp-model_annotated.html
- <http://www.life.uiuc.edu/crofts/bioph354/index.html>